

# Salinity and Nitrariaschoberi: Growth Parameters, Chlorophyll Content and Ion Accumulation

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**ABSTRACT:** Salinity is one of the most important factors that cause vegetation decline and sometimes elimination in arid and semi-arid regions. This study was carried out to investigate the effects of salinity on growth parameters, chlorophyll content and ion accumulation of Nitaria schoberi. Twenty pots were used in four replications and treatments consisted of a control treatment and four treatments with different salinity levels in a completely randomized design. Salt was added to the pots at concentrations of 0, 86, 160, 204 and 246 mM NaCI. Data analysis was performed by SPSS software. The first treatment (T1, 86 mM NaCI) achieved the best growth rate and the highest chlorophyll content and also the least amount of pigment degradation. According to the results, the ion balance of N. schoberi showed specific mechanisms in the tissues, so that the concentration of sodium and chloride ions increased with increasing salinity level. Best performance of Nitraria schoberi was obtained in low salinity (86 mM NaCI).

Keywords: Chlorophyll content, Growth Parameters, Ion Accumulation, Nitrariaschoberi, Salinity.

#### INTRODUCTION

Deterioration of biophysical environment of land by human-induced process resulting in reduced soil productivity is called land degradation (Dregne, 2002). The main causes of land degradation are (1) intensive cultivation, (2) deforestation, (3) poor quality irrigation, (4) unbalanced fertilization and (5) industrialization(Maqsood et al., 2013). According to the land degradation assessment in drylands (LADA, 2010), project executed by FAO, apart from the increased cost of fertilizer, land degradation is costing about 40 billion US dollars annually.

Soil salinity refers to the situation in which the presence of salts renders the soil to be of marginal quality. This leads to deflocculation/dispersion of soil colloids, translocated in the soil profile, blocks the conducting pores and reduces soil porosity (Vlek's et al., 2008).Salt affected soils impact vegetation through adverse physiological effects, and high levels of sodium degrade the physical structure of soil. Soil salinity results in the direct inhibition of plant growth or function due to osmotic stress, ion toxicity, or decreased absorption of essential nutrients (Hameed et al., 2008). However, plant species differ in their sensitivity or tolerance to salts (Brady and Weil, 1996). One of the most important properties of halophytes is their salinity tolerance (Liethet al.,1999). Nitraria schoberi is a halophyte, belongs to Zygophyllaceae family and toleratethe high salt concentrations (Mojiri and Jalilian, 2011). This species grows in the desert and alluvial floodplains with deep and sedimentary salty soils that have variable textures and calcareous and gypsic materials on the limited drainage conditions (Jori and Mahdavi, 2010).

Mojiri and Jalalian (2011) studied the relationship between the growth parameters of Nitraria schoberi and some soil properties. They concluded that the electrical conductivity and sodium ions had negative effects on plant growth and physiologic parameters.Panahi (2012) studied Salsolaarbuscula, S. orientalisand S. tomentosaresponses to salt stress and concluded that the rate of growth parameters increased with an increase in moderate salinity while in high salinity levels, salt stress caused all growth characteristics decline. Investigating halophytes response to salt with the aim of saline soils reclamation,Daoudet al. (2013) expressed that the salt-tolerant species survived at all salinity treatments, and the maximum growth occurred in low and moderate salinity (25 and 50% seawater).Brakez et al. (2013) studied the performance of Chenopodiumquinoa under salt stress and reported that the maximum biomass rate was recorded in 20% seawater treatment (low salinity level).

However, to face the increasing demands for food production, increased attention has been given to reclaim, improve and manage saline-sodic soil to solve salinity problems and achieve optimal crop production (sallam, 2007). The objective of this research is to investigate the effects of Salt Stress on growth, ion accumulation and phyto-biochemical parameters in Nitrariaschoberi.

#### METHODS AND MATERIALS

Experiment was conducted in the greenhouse of college of Natural Resources and Earth Sciences, University of Kashan located in Aran and Bidgul city, Kashan, Isfahan, Iran. Seeds of N. schoberi, from its natural habitat at Isfahan province, were seeded in plastic pots containing sand. After germinating and developing seeds in the seed bed for 3 months, the young seedlings were transplanted into 10 kg pots for a further 10 months (July 2012 through April 2013) with a standard dose of fertilizer (NPK) application.

For salinity treatments, twenty pots were used in four replications. Treatments were consisted of a control treatment and four treatments with different salinity levels in a completely randomized design. The NaCI-solutions were added to the pots at different concentrations of 0, 86, 160, 204 and 246 mM NaCl (Quiza et al., 2011).

After six weeks from the beginning of salt stress in all treatments, all planted seedlings were harvested. Leaves, stems and roots were weighed individually to determine fresh weight and oven dried at 80°C for at least 24 h. After drying, leaves, stems and roots dry weight was determined using a digital scale (AND, FX-300 GD, Japan) with an accuracy of 0.001 g.

Photosynthetic pigments (chlorophyll a and b) were estimated in the fresh plant tissue, after extracting with 85% acetone by Spectrophotometry method (by Spectrophotometer, Model 2100-UV) as described by Metzner et al. (1965). Wavelengths of 453, 505, 645 and 663 nm were chosen to measure these parameters by spectrophotometer. The parameters were calculated using the following equations:

- chlorophyll a  $\left(\frac{\text{mg}}{100 \text{ mL}}\right) = 0.999\text{A}663 0.0989\text{A}645$ chlorophyll b  $\left(\frac{\text{mg}}{100 \text{ mL}}\right) = -0.328\text{A}663 + 1.77\text{A}645$ Pigment degradation =  $1 \left(\frac{\text{total chl in salt treatment}}{\text{total chl in control}}\right) \times 100$ (Equation 1)
- (Equation 2)

(Equation 3)

Ash was prepared using Wilson's (1983) method. The extract obtained was used for the measurement of mineral elements. Chlorine measurement was performed using Mohr's method (1969). This volumetric method based on two sediments with quite distinct color of silver chloride and chromate. Flame photometrywas used to measure sodium and potassium ions accumulation rate (Portlab, Model 378P, Italy). Calcium and magnesium were obtained by titration with EDTA 0.01N.

Statistical analyses were doneby mean comparison of Duncan's multiple range method using SPSS software version 16.0.

#### **RESULTS AND DISCUSSION**

## Growth parameters

According to the results of analysis of variance, root and leaf dry weight along with the fresh and dry weight of the total plant showed a significant (P<0.05) difference between different treatments (Table 1). At the end of the experiment, 75% of the forth treatment (T4) (246 mM NaCl) was almost dried. The first treatment (T1) (86 mM NaCl) with a salinity level equal to 0.1% of the pots dry weight (86 mM NaCl) had the highest rates in all obtained data from the different organs of the plant. It was confirmed with comparison of data (average dry weight of roots, stems and leaves along with a total biomass in different treatments) and it can be said that T1 has more growth in comparison to the others (Figure 1). These results coincide with those obtained by Panahi (2012) studying Salsolaorientalis, Daoud et al. (2013) in the investigation of halophytes response to salt and Brakezet al. (2013) studying Chenopodiumquinoa. They also expressed that maximum growth occurred in low and moderate salinity. As a result, the salinity decrease efficiency of the roots in providing nutrients and water to other organs declines shoot growth, reduces the organogenesis and production of dry matter, leads to reduced nutrient transport and ultimately leads to a reduction in root and stem weight (Daoudet al., 2013). The decline in growth as a result of high soil salinity is mainly attributed to osmotic inhibition of water uptake, accumulation of ions such as Na<sup>+</sup> or Cl<sup>-</sup> in pant cells and insufficient uptake of essential nutrients (Munns and Tormaat, 1986). The response of the plants to different levels of salinity was traduced at the scale of the whole plant by the simulation of biomass, expressed by production of dry matter, at low and moderate salinities and a progressive decrease of biomass weight in high salinity treatments. It is a typical reaction of halophytes to saline stress (Kelley et al., 1982; Glenn and O'Leary,

1984; Munns and Termatt, 1986; Mass, 1987; Gorham, 1996; Munns, 2002; in Daoudet al., 2013). Plant's root has importance in plants survival in the presence of salts due to the ability of the root system to control the entry of ions to shoots (Hajibagheri et al., 1989), and the lack of water uptake by plants due to high salinity causes reduced shoot and root growth(Werner and Finkelstein, 1995).

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S.O.V	df	M.Ś								
		Root FW	Stem FW	Leaf FW	Fresh biomass	Root DW	Stem DW	Leaf DW	Dry biomass	
Salinity	4	18.318	3.936 <sup>ns</sup>	14.565 <sup>ns</sup>	91.419	7.937	0.926 <sup>ns</sup>	0.999	19.754	
Error	15	5.736	1.573	4.821	26.122	1.951	0.511	0.226	5.021	



Figure 1.Salinity effects on growth parameters in Nitrariaschoberi

## Chlorophyll content

Based on the analysis of variance, chlorophyll "a" and total chlorophyll showed a very significant relationship (P<0.01) with increasing salinity (Table 2). Chlorophyll "b" also showed a significant relationship (P<0.05) with increasing salinity. Duncan test results also showed that chlorophyll "a" increased with increasing salinity levels to first treatment (T1, 86 mMNaCl) butdecreased with further increases in salinity levels (Figure 2.1). According to the analysis of variance, pigment degradation has showed a very significant relationship (P<0.01) with increasing salinity levels (Table 2). Results of Duncan test also showed that pigment degradation decreased with increasing salinity levels to first treatment (T1, 86 mMNaCl) and increased with further increases in salinity levels (Figure 2.4). These results were confirmed by Abbasi et al. (2009) studying Rubia tinctorum. They concluded that chlorophyll content increased up to 15dS.m<sup>-1</sup> and then decreased with increasing salinity levels.

Table2.	Results of analys	sis of variance for the impa	act of NaCl stress or	n chlorophyll contents	in N. schoberi.			
		M.S						
S.O.V	df	chlorophyll a	chlorophyll b	total chlorophyll	pigment degradation			
salinity	4	0.017	0.005	0.038	4199.903**			
error	15	0.002	0.001	0.004	433.685			
		"Significant in P<0.	01, <sup>*</sup> Significant in P	<0.05.				



Figure 2. Effects of salinity on chlorophyll content in Nitrariaschoberi.

It is possible that in high concentrations of salts, the binding of chlorophyll and chloroplast proteins are loose and chlorophyll is destroyed due to the effect of ions on proteins (Abbasi et al.,2009). Chlorophyll contentincrease can be partly attributed to leaf thickness increase in saline environments (Abbasi et al.,2009).

#### lon accumulation Sodium (Na<sup>+</sup>)

Analysis of variance revealed a significant effect (P<0.05) of salinity on the concentration of sodium ion in different plant organs in N.schoberi(Table 3, Figure 3). According to the comparison of different levels of salinity, it can be concluded that a strong and direct relationship exists between increasing salinity and content of sodium ions in different plant organs (Figure 3). Through various organs, leaves have the highest sodium rates and roots have the lowest rate. So that the highest concentration of sodium ions in different organs (root, stem and leaf) was observed in the highest level of salinity (T4, 246 mM NaCl) (22856.78, 25840.02 and 131250.00ppm respectively) and the lowest values were recorded in control (15797.22, 17286.12 and 67500.00ppm, respectively). Also among the measured ions, sodium had one of the highest values. With this regard, the greatest accumulation of sodium by plants at high salt concentration may be attributed to the damage of plant cells protoplasm and, as a result of the selective salt absorption, is replaced by passive absorption which causes abnormal accumulation of salts in plant organs (Kader and Lindberg, 2005). Khan et al. (2009) proved that specific ions such as sodium may have toxic effects on plants: reducing growth or causing damage to cells and membranes. These results coincide with those obtained by Daoud et al. (2013) in the investigation of halophytes response to salt, Brakezet al. (2013) studying

Chenopodiumquinoa and Tawfik et al. (2013) studying Leptochloafusca and Sporobolusvirginicus. They expressed that Na<sup>+</sup> concentration increased with increasing salinity levels.

S.O.V salinity error M.S df 4 15 1.117×10<sup>4</sup> Root 3.439×10<sup>4</sup> Sodium Stem 4.323×10<sup>7</sup>  $1.299 \times 10^{7}$ 3.160×10<sup>9\*\*</sup>  $8.890 \times 10^{7}$ Leaf 3.401×10<sup>7\*\*</sup> 1.136×10<sup>8</sup> Root 1.601×10<sup>9\*\*</sup> 8.208×10<sup>6</sup> Chloride Stem 1.675×10<sup>9\*</sup> 4.102×10<sup>6</sup> Leaf Root  $1.129 \times 10^{8}$  $3.111 \times 10^{7}$ 1.695×10<sup>7ns</sup> Potassium Stem  $1.003 \times 10^{7}$ 2.638×10<sup>8\*</sup> Leaf  $2.366 \times 10^7$ 7.951×10<sup>8\*</sup> Root 2.013×10<sup>7</sup> 1.636×10<sup>7ns</sup>  $4.388 \times 10^{7}$ Calcium Stem 5.228×10<sup>8\*\*</sup> 7.264×10<sup>8</sup> Leaf 6.688×10<sup>8\*\*</sup>  $1.245 \times 10^{8}$ Root 3.014×10<sup>7ns</sup> Magnesium  $1.149 \times 10^{7}$ Stem 3.679×10<sup>8\*\*</sup> leaf 2.207×10<sup>7</sup>

Table3. Results of analysis of variance for the impact of NaCl stress on ions accumulation in Nitrariaschoberi.

Significant in P<0.01, Significant in P<0.05, ns non-significant



Figure 3.Effects of salinity on sodium content in Nitrariaschoberi.

## Chloride (Cſ)

The results well confirm the influence of salinity on chloride concentrations in various organs of N.schoberi and causes a very significant increase (P<0.01) in the concentration of chloride (Figure 4, Table 3). Statistical comparison shows that the chloride concentration of various organs increased directly with increasing salinity. Treatment 4 (246 mM NaCl) has obtained the highest rate of chloride concentration (23193.33, 66473.75 and 146881.30ppm in root, stem and leaf, respectively) and control has showed the lowest rate of chloride accumulation (16063.75, 22365.00 and 96156.19ppm, respectively) (Figure 4). The chloride accumulation of leaf was higher than other tissues. Flowers et al. (1977) and Greenway and Munns (1980) (in Koyroet al., 2008) reported that halophytes need an excess of salt for maximum growth and for maintaining low solute potentials. The accumulation of NaCl consumes much less energy than needed for the osmotic adjustment with compatible organic solutes (Yeo, 1983). These results coincide with those obtained by Daoud et al. (2013) in the investigation of halophytes response to salt stress in which Cl concentration increased with increasing salinity levels.



Figure 4.Effects of salinity on chloride content in Nitrariaschoberi.

# Potassium (K<sup>+</sup>)

Results indicate a significant inverse effect of salinity on potassium ions concentration in different organs of N. schoberi. Comparison results indicate that the concentrations of potassium in leaves of this species increased very significantly (P<0.01) up to the first treatment (86 mM NaCl) and then decreased with increasing salinity levels (Figure 5, Table 3). Concentrations of potassium in roots also increased significantly (P<0.05) up to first treatment (86 mM NaCl) and then decreased with increasing salinity levels (Figure 5, Table 3). Concentrations of potassium in roots also increased significantly (P<0.05) up to first treatment (86 mM NaCl) and then decreased with increasing salinity levels (Figure 5, Table 3). Potassium concentration in the stem didn't show a significant difference in different treatments (Table 3). Na<sup>+</sup> concentration affects K<sup>+</sup> and Ca<sup>2+</sup> absorption, which were in low concentrations compared to Na<sup>+</sup> (Brakezet al., 2013). Potassium is considered as one of the primary osmotic substances which contribute to osmotic adjustment in many plant species (Ashraf et al., 2001). Higher potassium levels in plants are beneficial in achieving a better plant survival with improved plant growth under saline stress (Sangakkaraet al., 2001; Umar, 2002). These results coincide with those obtained by Daoud et al. (2013) in the investigation of halophytes response to salt stress and Tawfiketal. (2013) studying Leptochloafusca and Sporobolusvirginicus. They declared that K<sup>+</sup> concentration decreased with increasing salinity levels.



Figure 5.Effects of salinity on potassium content in Nitrariaschoberi.

# Calcium (Ca<sup>2+</sup>)

Results indicate a significant inverse effect of salinity on calcium ions concentration in different organs of N.schoberi. Comparison results indicate that calcium concentration in the leaves of this species increased very significantly (P<0.01) up to first treatment (86 mM NaCl) and then decreased with increasing salinity levels (Figure 6, Table 3). Calcium Concentration of roots also increased significantly (P<0.05) up to first treatment (86 mM NaCl) and then decreased with increasing salinity levels (Figure 6, Table 3). Calcium Concentration of roots also increased significantly (P<0.05) up to first treatment (86 mM NaCl) and then decreased with increasing salinity levels (Figure 6). Calcium concentration of stem didn't show a significant difference in different treatments. The accumulation of calcium in root was higher than other tissues. The

Na<sup>+</sup> ion uptake by plant limits the absorption of essential ions such as K<sup>+</sup> and Ca<sup>2+</sup> (Jendoubi, 1997). The Ca<sup>2+</sup> increased salt tolerance of Plants (Munns et al. 1983; Epstein 1998) by its function in maintaining membrane integrity and controlling the selectivity of plant ion uptake and transport under saline conditions (Lauchli and Wieneke, 1979; Lynch and Lauchli, 1985; Marschner, 1995). Calcium plays a vital role in maintaining membrane stability and permeability (Mengel and Kirkby, 1987). The Mg<sup>2+</sup> is an important element occupying a central position in the chlorophyll molecule, thereby affecting the concentration of chlorophylls "a" and "b" in the green plant tissues (Yasseen and Al-Thani, 2007). The results of this study coincide with those obtained by Daoud et al. (2013) in the investigation of Spartina alterniflora and Tawfik et al. (2013) studying Leptochloafusca and Sporobolusvirginicus. They expressed that Ca<sup>2+</sup> concentration decreased with increasing salinity levels.



Figure 6.Effects of salinity on calcium content in Nitrariaschoberi.

## Magnesium (Mg<sup>2+</sup>)

Salinity levels that used in the present study affected very significantly (P<0.01) on the accumulation of magnesium in leaves and roots of N. schoberi (Table 3). Although there was no significant effect of salinity on the rate of this parameter in the stems. The Statistical comparison suggests that magnesium accumulation in leaves and roots significantly (P<0.01) increased with increasing salinity levels compared with control, however, no significant differences were observed between the different levels of salinity (Figure 7). StudyingLeptochloafusca and SporobolusvirginicusTawfiket al. (2013) noted to no effect of salinity increase in Mg<sup>2+</sup> concentration. The results of the statistical comparison of means shows that the effects of salinity on the magnesium ion concentration in the roots are more than stems and leaves (Figure 7). The Mg<sup>2+</sup> is an essential constituent of chlorophyll and some glycolysis enzymes, its concentration increased with increasing water salinity concentration in the culture medium, for this reason plants did not show any chlorosis symptoms due to Mg<sup>2+</sup> deficiency up to treatment 4 (246 mM NaCl). The Mg<sup>2+</sup> is an important element occupying a central position in the chlorophyll molecule, thereby affecting the concentration of chlorophylls "a" and "b" in the green plant tissues (Yasseen and Al-Thani, 2007).



Figure 7.Effects of salinity on magnesium content in Nitrariaschoberi.

#### Relationship between ions

Increasing salinity, the ratio of potassium ions to sodium ions were significantly decreased and treatment 1 (86 mMNaCl) had the highest value (Figure 8.1, Table 4). There is no significant difference between treatment 1 and control (Figure 8.1). A similar trend is seen in the ratio of calcium ions to sodium ions (Figure 8.2, Table 4). Treatment 1 is achieved the highest ratio of magnesium ion to sodium ion (Figure 8.3). Besides the high relative water content (Ghorbaniet al., 2014), Ca<sup>2+</sup>/Na<sup>+</sup> ratio increase (in treatment 1, 86 mM NaCl) has supported these results (best performance of N. schoberi in the first treatment). These conditions prevent Ca<sup>2+</sup> deficiency induced by Na<sup>+</sup> and improve growth stimulation of plants (Marschner, 1995). The survival of halophytes in high Na<sup>+</sup> concentrations conditions depends on their capacity to maintain low Na<sup>+</sup> concentrations and high K<sup>+</sup>/Na<sup>+</sup> in the cytoplasm level (Blumwaldet al., 2000). Exclusion of Na<sup>+</sup> (in salt excluders) and/or its compartmentation in the vacuole for osmotic role (in salt includers) are strategies to maintain a high K<sup>+</sup>/Na<sup>+</sup> (Greenway and Munns, 1980; Blumwaldet al., 2000). Potassium is considered as one of the primary osmotic substances which contribute to osmotic adjustment in many plant species (Ashraf et al., 2001). Sustained K<sup>+</sup>/Na<sup>+</sup> selectivity is therefore proposed as a physiological marker for the ionic component of salt stress, providing ion homeostasis in growing roots (Muhling and Lauchli, 2001).





Figure 8.The effect of salinity on the relationship ofions in Nitrariaschoberi.

In order to mediate osmotic adjustment, achieve low water potential gradient from substrate to different parts of the plants, protect subcellular structures and reduce oxidative damage caused by free radicals that are produced in response to high salinity, compatible solutes, especially in salt excluding species, and at lower degree for salt including ones, are synthesizes by plants as a metabolic response to salt stress (Hong et al., 1992; Hare et al., 1998). The synthesis of organic solutes, which can be alcohols; soluble carbohydrates or organic acids, is energy demanding (Luttge, 2002; Koyroet al., 2008) and the formation of these solutes deceases the energy status of the plant and can lead to decreases of growth in high saline condition (Koyro, 1997; Koyro and Huchzermeyer, 1999a). Lacerda et al. (2005) reported that the greatest salinity tolerance observed in plants under saline conditions was associated with higher K<sup>+</sup>/Na<sup>+</sup> ratio and greater capacity for osmotic adjustment.

#### CONCLUSIONS

Looking over the data, it is clear that the studied species is capable of tolerating salinity. Best performance of Nitraria schoberi was obtained in low salinity (86 mM NaCl). However, increasing salinity more than the optimal rate, the growth efficiency of plant decreased. Finally, it can be stated that N.schoberi is suitable for saline

conditions and capable of growing in such detrimental conditions, but further investigations are needed to evaluate its performance in different conditions and under multi-stress situations.

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